

Prevalence and Concentration of *Cryptosporidium* Oocysts in Beef Cattle Paddock Soils and Forage

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Abstract

Cryptosporidium is a parasitic protozoan of great interest because of its widespread occurrence in surface waters, high degree of infectivity, and difficulty of risk management associated with its presence and control. Information about environmental loading and seasonal prevalence of *Cryptosporidium* oocysts is important for development of watershed management plans to protect public health. Healthy adult beef cattle are known to shed oocysts into the environment, and *Cryptosporidium* oocysts are often present all year in streams and groundwater in livestock agriculture areas. Surface soil and forage samples from 12 Virginia, United States, paddocks were analyzed bimonthly over 3 years for the presence and concentrations of *Cryptosporidium* oocysts. Half of the paddocks were grazed by stocker beef from November to September. The other half were managed for hay, but were grazed for a few days by the same animals in late fall and early spring to clean up late fall forage regrowth. Annual mean oocyst prevalences in soil were 57.9% and 48.4% in pasture and hay paddocks, respectively. Mean annual oocyst prevalences on forage were 52.4% and 40.5% in pasture and hay paddocks, respectively. Prevalence and concentration of oocysts on hay forage was highest in summer. Oocyst concentrations increased with increasing prevalences in both management systems. Wild animals appeared to be efficient vectors for oocyst distribution among paddocks. Canopy management, short-cycle rotational grazing, and control of wildlife are potential strategies for reduction of *Cryptosporidium* oocysts in pasture and lessening risk of contamination of water supplies, but further studies are needed before recommendations can be made.

Introduction

CRYPTOSPORIDIUM PARVUM, an enteric coccidian protozoan that is an emergent zoonotic pathogen, has received an increasing amount of study interest because of its widespread occurrence in surface waters (LeChevallier *et al.*, 1991), its high degree of infectivity (Dupont *et al.*, 1995), and the difficulty of risk management associated with its presence and control (Hamilton *et al.*, 2006). Karanis *et al.* (2007) highlighted three life cycle features of *Cryptosporidium* that enhance the likelihood of waterborne transmission: monoxenous nature, zoonotic transmission, and environmental robustness. Transmission of the oocysts between animals and humans occurs by the fecal/oral route or by ingestion of contaminated food or water (King and Monis, 2006). Cryptosporidiosis is a disease caused by ingestion of *C. parvum* oocysts that have been excreted in feces of infected humans or animals. Ingestion of small numbers of oocysts can cause infection (CDC, 1995; Dupont *et al.*, 1995). Disease symptoms develop within 2–10 days of ingesting *C. parvum* oocysts and include diarrhea that lasts 1–2 weeks, abdominal cramps, fatigue, nausea,

vomiting, and low-grade fever. Cryptosporidiosis can be chronic and life threatening to persons with weakened and compromised immune systems.

Transport of *C. parvum* oocysts to water sources is not completely understood. Agricultural sources of oocysts can take three different routes to water supplies. Oocysts are sometimes deposited directly in water or near-stream source areas by deposition of feces by grazing animals or application of manure for soil improvement (Hubbard *et al.*, 2004; Schijven *et al.*, 2004). Overland runoff transports oocysts to water bodies (Davies *et al.*, 2004) and oocysts are transported by infiltrating water (Mawdsley *et al.*, 1995), especially in macroporous soils (Harter *et al.*, 2008; Boyer *et al.*, 2009).

Information about total number of oocysts deposited on soil and plant surfaces (environmental loading) and seasonal prevalence of oocysts is important for development of watershed management plans to protect public health (Atwill *et al.*, 2006). Calves are usually considered to be the primary source of oocysts (up to 10^7 oocysts/g of manure) in beef production (Atwill *et al.*, 1999), but apparently healthy adult beef cattle have been shown to shed oocysts in concentrations of 25 to

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1.8×10^4 oocysts/g of manure (Scott *et al.*, 1994). Studies have found *C. parvum* oocysts present all year in streams flowing through livestock agriculture areas, but highest frequency of oocyst occurrence coincides with calving season (Kemp *et al.*, 1995; Ong *et al.*, 1996; States *et al.*, 1997; Bodley-Tickell *et al.*, 2002). Boyer and Kuczynska (2003) found oocyst concentrations in a karst spring to be greatest in late fall to early winter even though calving season was in the early spring. They concluded that oocysts were transported to epikarst where oocysts accumulated until saturated hydrologic conditions flushed oocysts into karst conduits and subsequently to springs.

On-farm strategies to reduce foodborne pathogens depend on an understanding of a myriad of complex interactions (Callaway and Oliver, 2009). Development of grazing strategies and pasture management systems that minimize opportunities for *C. parvum* oocysts to reach drinking water sources requires basic knowledge about temporal and spatial distributions of oocyst deposition densities on landscapes. Armed with that basic knowledge, pasture and livestock managers have the opportunity to develop management systems that reduce environmental loading of oocysts, increase the opportunity for environmental conditions to deactivate oocysts, and keep oocysts out of runoff source areas. The purpose of this study was to investigate the temporal distribution of oocysts at the soil surface and on standing forage in rotationally grazed beef pastures.

Materials and Methods

The study was conducted on a site consisting of Angus-cross stocker cattle grazing tall fescue (*Schedonorus phoenix*)

pasture and fed fescue hay as needed. The pasture was located at Virginia Tech's Shenandoah Valley Agricultural Research and Extension Center, Steele's Tavern, VA (37°55.77' N, 79°13.27' W; 540–552 m above sea level) (Fig. 1). Runoff from the road and hill above the road were diverted away from the study site by roadside ditches. Study weather data were obtained from the U.S. Weather Bureau station at Lexington, VA, located about 25 km south of the study site. Soils at the study site are primarily Frederick (fine, mixed, semi-active, mesic Typic Paleudults) and Carbo (very-fine, mixed, active, mesic Typic Hapludalfs) silt loams. The study site is located on karstified carbonates consisting of Ordovician limestone and dolostone.

The 14 ha pasture was divided into 24 paddocks (~0.6 ha each) that were rotationally grazed by stocker beef cattle. Twelve of the paddocks were used for this study (Fig. 1). Poultry litter was used as a soil amendment in the 12 excluded paddocks. The 12 selected paddocks underwent typical management for the region. Sample sizes were determined by availability of laboratory and personnel resources. Three samples per paddock on each visit were considered representative. Even-numbered paddocks (hay paddocks) were used to grow hay for stockpile feeding cattle in the odd-numbered paddocks (pasture paddocks) during winter. Hay paddocks were fertilized in fall and hayed in summer; cattle were rotated in for cleanup grazing of fall forage regrowth before winter rest. Pasture paddocks were stocked with four 180 kg Angus-cross steers each in November. Steers were rotationally grazed in the pasture paddocks until the cattle were sold at a weight of about 400 kg in August or September of the following year and new stocker steers were purchased in

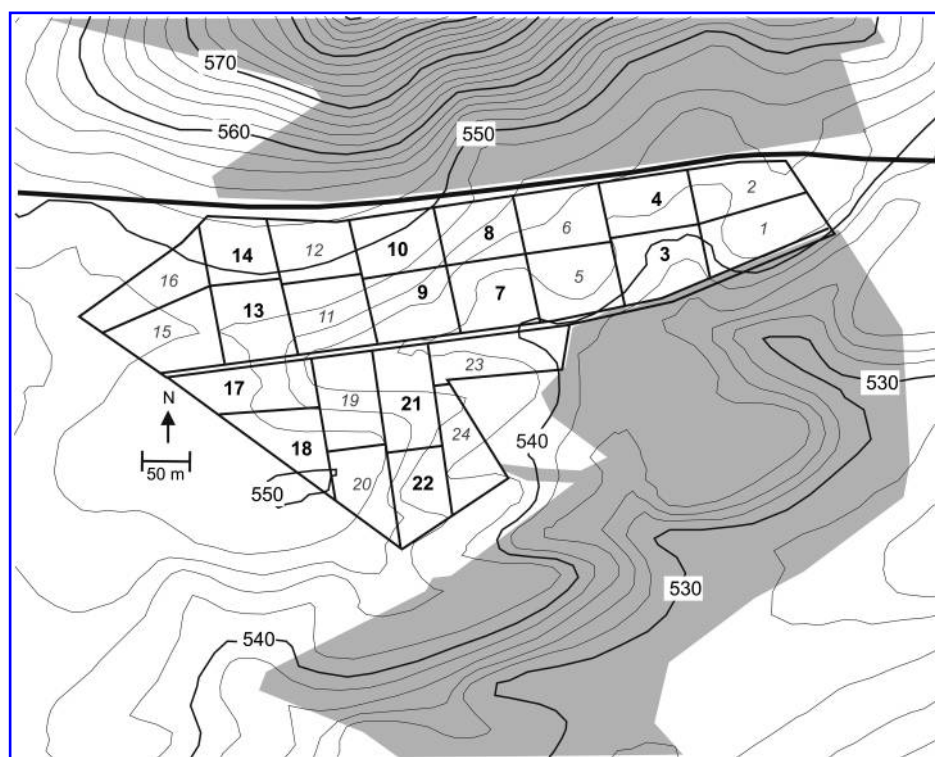


FIG. 1. Study site at Steele's Tavern, VA. Paddocks used in this study are numbered in bold lettering. Shading represents forest, and elevation contour interval equals 2 m.

November from Midwestern U.S. suppliers. High-energy supplemental feeds for the cattle included corn and broiler litter.

The paddocks were visited bimonthly from May 2002 to November 2004, and the top 2.5 cm of soil beneath the thatch was sampled with a 5-cm-diameter soil probe at three separate arbitrary locations in each paddock where vegetation cover was present. Samples were not obtained from the same points during subsequent visits. A bimonthly sampling strategy was employed because of the long travel distance and resource limitations. Soil samples were not taken in proximity (~1 m) to manure paddies or in low spots such as hoof prints. The three soil samples from each paddock were mixed by hand in sterile plastic bags and transported back to the laboratory for analyses of *Cryptosporidium* oocyst concentrations. Forage samples were also taken at each soil sample location, and combined and mixed by hand in sterile plastic bags in each paddock on seven of the sample months (May, July, December 2002; March, September, November 2003; May 2004) for *Cryptosporidium* oocyst analyses.

Soil (25 g) and grass (4 g) samples were processed using the NaCl flotation method as previously described (Kuczynska and Shelton, 1999). Three 10 μ L aliquots (of the final 100 μ L suspension) were pipetted onto slide wells (5 mm diameter), dried on a slide warmer, and stained using a commercial immunofluorescence antibody kit (Merifluor; Meridian Diagnostic, Cincinnati, OH). Numbers of oocysts per soil or grass sample were obtained by multiplying the average number of oocysts in three wells by 10. Microscope slides were examined with an epifluorescence microscope (Zeiss; Carl Zeiss MicroImaging, Thornwood, NY) at 250 \times magnification. No speciation or test for infectivity was done on the oocysts since we were only looking at oocysts as indicators of *Cryptosporidium*.

Oocyst concentrations were converted to number of oocysts per gram of soil or grass, and oocyst concentration statistics were calculated from all oocyst-positive samples. Statistical analyses were performed with SAS Version 9.1 (SAS Institute, Cary, NC). The LOGISTIC procedure was used to test for season or treatment effects on the presence or absence of oocysts. The presence/absence of oocysts was treated as a binary variable, where presence was coded as 1 and absence as 0. Deviance Chi-square (called -2 Log L in SAS) was used to test the goodness of fit of the logistic model. Sign tests using paired samples tested for differences in oocyst prevalences and concentrations. The paired samples were paddock or seasonal means. Repeated measures were not considered since sign tests were performed on means. Linear and curvilinear regression were used to test for relationships between mean oocyst prevalences and mean oocyst concentrations. Statistical tests are significant at $p \leq 0.1$ level unless stated otherwise.

Results

Figure 2 shows observed and normal total precipitation and mean daily temperature, by months, for the study years. Mean daily temperature was nearly normal all 3 years with the exception of cooler than normal temperatures during the 2003 growing season. The cooler than normal temperatures in 2003 were associated with a wetter than normal spring and summer.

Table 1 summarizes mean bimonthly and annual prevalences of oocysts in surface soils and on forage in pasture and

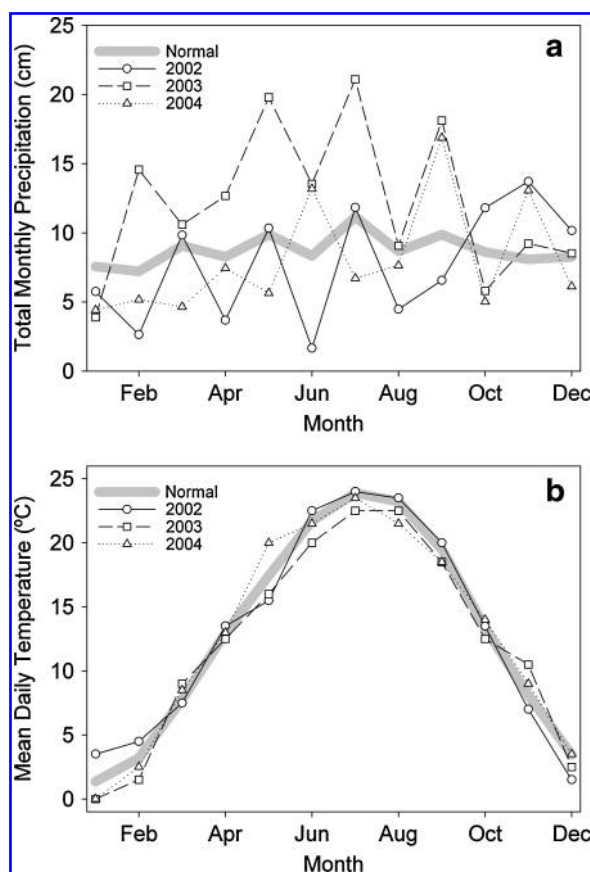


FIG. 2. Monthly and normal climatic conditions at Lexington, VA, for the years of study: (a) total monthly precipitation (cm); (b) mean daily temperature (°C).

hay paddocks. Table 1 also shows bimonthly and annual mean concentrations of oocysts per gram of soil or forage in samples that were positive for *Cryptosporidium* oocysts. Bimonthly prevalences of oocysts in pasture or hay soils were not significantly different. Bimonthly prevalences of oocysts on forage were significantly different in pasture and hay paddocks. Bimonthly prevalences of oocysts on forage were lowest in winter to early spring and highest in summer to early fall.

Mean bimonthly concentrations of oocysts in surface soil samples that tested positive for oocysts ranged from 0.17 to 0.27 oocysts/g in the pasture paddocks and 0.16 to 0.22 oocysts/g in the hay paddocks. Mean bimonthly concentrations of oocysts in soil were not significantly different in the pasture or the hay paddocks. Mean bimonthly concentrations of oocysts on forage samples that tested positive for oocysts were significantly different in the hay paddocks, but not in the pasture paddocks. Mean bimonthly concentrations of oocysts on forage in the hay paddocks ranged from 0.81 oocyst/g in May and November to 1.56 oocysts/g in September. Although mean bimonthly oocyst concentrations on forage in pasture were not significantly different, they ranged from 0.81 to 2.31 oocysts/g.

In the 16 months sampled, prevalence of oocysts was greater in the pasture paddock soils than in the hay paddock soils eight times. Prevalence of oocysts in soil was equal in both paddock treatments in three of the sample months. No

TABLE 1. SUMMARY OF THE MEAN BIMONTHLY AND ANNUAL PREVALENCES AND CONCENTRATIONS OF *CRYPTOSPORIDIUM* OOCYSTS IN POSITIVE SAMPLES OF SURFACE SOIL AND FORAGE IN THE PASTURE AND HAY PADDOCKS

Month	Pasture				Hay			
	Soil		Forage		Soil		Forage	
	Prevalence (%)	Mean oocysts (no./g)	Prevalence (%)	Mean oocysts (no./g)	Prevalence (%)	Mean oocysts (no./g)	Prevalence (%)	Mean oocysts (no./g)
Jan	50.0	0.21	n.d.	n.d.	63.6	0.16	n.d.	n.d.
Mar	66.7	0.19	16.7	0.81	50.0	0.20	0.00	n.d.
May	52.9	0.18	50.0	1.25	33.3	0.22	33.3	0.81
July	72.2	0.25	66.7	2.31	50.0	0.19	66.7	1.13
Sep	66.7	0.27	66.7	1.56	66.7	0.18	66.7	1.56
Nov	38.9	0.17	58.3	1.25	33.3	0.17	41.7	0.81
Year	57.9	0.22	52.4	1.44	48.4	0.18	40.5	1.00

Statistically significant differences in seasonal means are indicated by italicized numbers in a column. Differences between pasture and hay are indicated by asterisks at the top of the columns (i.e., two columns with the same number of asterisks are significantly different). n.d., not determined.

significant difference between bi-monthly prevalences of oocysts in pasture versus hay paddock soils was observed.

Oocyst concentrations in pasture paddock soils were greater than oocyst concentrations in hay paddock soils when considering bi-monthly means. Most of the difference occurred in summer and fall when oocyst concentrations were 0.06 to 0.09 oocysts/g of soil greater in the pasture paddocks (Table 1). Mean oocyst concentrations on the forages also differed. Oocyst concentrations on pasture paddock forages were about twice the concentration (2.31 vs. 1.13 oocysts/g) of oocysts on hay paddock forages in July.

Seasonal shifts in oocyst prevalence and concentration on pasture forage suggest that there might be a relationship with air temperature. Figure 3 shows trends between mean monthly air temperature and mean prevalences and mean concentrations of oocysts on the pasture forage. Although the bimonthly concentrations of oocysts on pasture forage were

not significantly different, the concentrations did follow the temperature pattern. Warmer air temperatures might change oocyst and vegetation surface properties, thereby enhancing the ability of oocysts to cling to vegetation.

Direct relationships between oocyst prevalence and concentration have been observed on an English lowland farm (Sturdee *et al.*, 2003) and in an upland watershed in England (Sturdee *et al.*, 2007). A linear relationship was observed for oocyst concentration versus prevalence on pasture forage in this study (Fig. 4). Mean oocyst concentrations on pasture forage increased about 0.32 oocysts/g for each 10% increase in prevalence. An increase in oocyst concentration with increased oocyst prevalence in pasture soil was also observed with mean concentration increasing about 0.035 oocysts/g for each 10% increase in prevalence (Fig. 5). All of the lines in Figures 4 and 5 were forced through the origin by using regression analysis with no intercept, because the concentration of oocysts has to be zero if prevalence of oocysts is zero. Obviously, if an intercept term was included in the regression

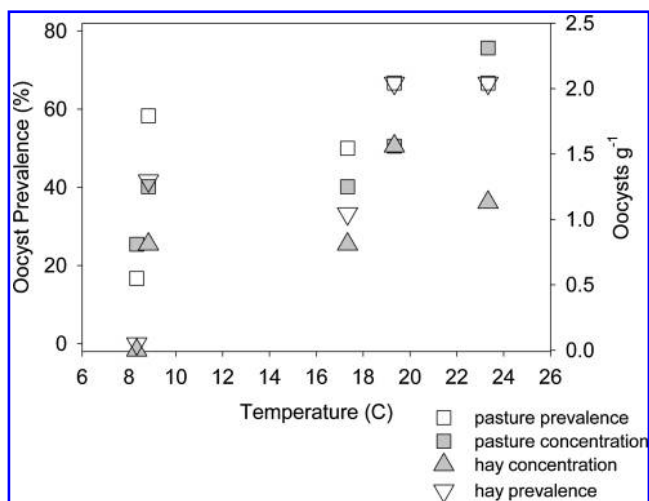


FIG. 3. Mean oocyst prevalences (open symbols) and mean oocyst concentrations (filled symbols) versus mean monthly temperature on pasture (squares) and hay (triangles) paddock forages.

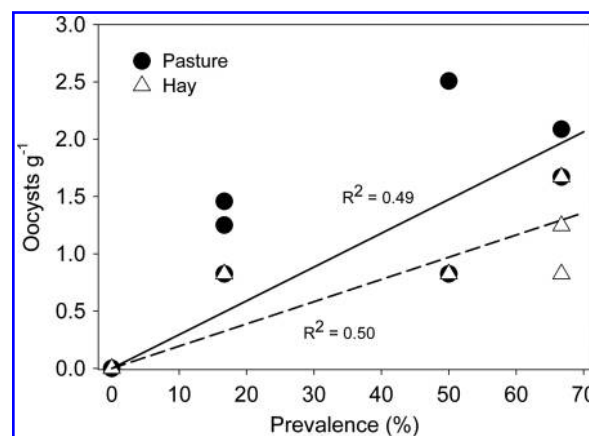


FIG. 4. Mean oocyst concentrations versus mean oocyst prevalences on pasture paddock (filled circles) and hay paddock (open triangles) forages. Regression lines represent trends for pasture (solid line) and hay (dashed line). Regression analyses were run with the no intercept option.

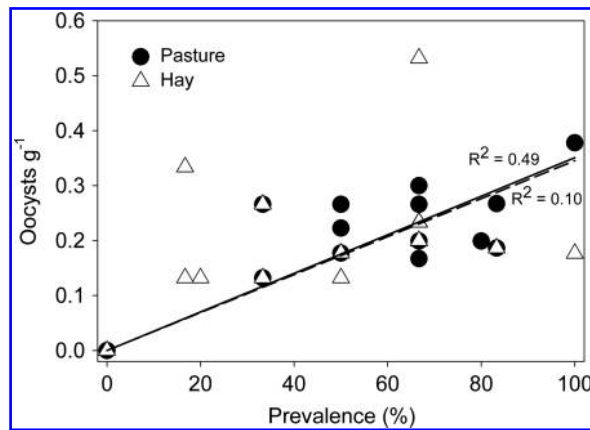


FIG. 5. Mean oocyst concentrations versus mean oocyst prevalences in pasture paddock (filled circles) and hay paddock (open triangles) soils. Regression lines represent trends for pasture (solid line) and hay (dashed line). The slope of the regression line for hay is not statistically different from zero. Regression analyses were run with the no intercept option.

analyses, the R -squares would be lower. Curvilinear line fitting did not improve predictability associated with the relationships.

No significant differences were found in the annual prevalences of oocysts. There were significant differences in the annual mean concentrations of oocysts. Annual mean concentrations of oocysts were 0.04 oocysts/g greater in pasture paddock soils than hay paddock soils. Annual mean concentrations were 0.44 oocysts/g greater on pasture paddock forages than hay paddock forages.

Discussion

Pasture paddocks

Uniformity of oocyst prevalences and concentrations throughout the year in pasture soils suggests that oocysts were being deposited year round. Further, oocyst deposition rates had to be equal to rates of oocyst removal to maintain the uniformity in prevalences and concentrations of oocysts in the surface soil. Oocysts could have been removed from pasture soils by surface runoff, subsurface transport with infiltrating water, and die-off with subsequent microbial decomposition. Studies have shown that rainsplash can relocate fecal bacteria (Boyer and Belesky, 2009) and fungal spores (Paul *et al.*, 2002) from soil forage and grass surfaces. A similar mechanism of oocyst removal from pasture soils would be expected.

Prevalence and concentration of oocysts were expected to be highest in late fall when younger stocker cattle were present. *Cryptosporidium* infection is known to be more prevalent in young calves than in adult cattle. Infections of adult cattle by *Cryptosporidium* are known to occur, but with excretion of lower concentrations of oocysts (Fayer *et al.*, 2000). The greater quantity of manure deposition by adult cattle than young stocker cattle could account for a uniform environmental loading of oocysts throughout the year (Kuczynska *et al.*, 2005). Barwick *et al.* (2003b) found that 17% of soil samples from New York dairy farms tested positive for *Cryptosporidium* and that 92% of farms had at least one soil

sample test positive for *Cryptosporidium*. The high farm prevalence might have been a result of accumulation of feces in soil over time (Barwick *et al.*, 2003a). Oocysts were more likely to be found in soil of land areas that accumulated fresh manure over time (Barwick *et al.*, 2003b).

Bimonthly oocyst prevalences on the pasture forage changed systematically with the lowest prevalence in winter and the highest in summer. Oocyst concentrations on pasture forage did not show significant bimonthly changes even though mean concentrations followed a pattern similar to bimonthly prevalences. In summer, cattle roam throughout paddocks in search of food and would be expected to excrete manure more evenly over paddocks as opposed to winter, when most of the feeding is done with supplemental hay. By concentrating cattle near hay feeders, cattle do not roam the paddocks in search of food, and manure deposition is also concentrated near hay feeding stations.

Hay paddocks

Similar to the pasture paddocks, bimonthly oocyst prevalences and oocyst concentrations in hay paddock soils did not change significantly across the year. That was surprising since cattle were rarely present in the hay paddocks. Cattle were generally present in the hay paddocks in early spring before significant new grass growth occurred and late summer following the last hay harvest and before the fall sale of cattle. Mean oocyst prevalence in September was highest observed mean (66.7%), but it was not significantly different from those in other months (Table 1).

Bimonthly oocyst prevalences on the hay paddock forage followed a clear trend with highest prevalences and concentrations in late summer and early autumn and lowest in winter (Table 1). The trend was expected given that cattle were present in hay paddocks at times of highest oocyst prevalences and concentrations. In the period from the time cattle were sold (September) to the time just before new stocker cattle were introduced (November), the oocyst concentrations on forage dropped nearly 50% from 1.56 to 0.81 oocysts/g of forage. At the same time, prevalence of oocysts on the hay paddock forages dropped about 37%.

Mean oocyst prevalences and concentrations on hay paddock forages appeared to be related to mean monthly temperature (Fig. 3), indicating that there was some seasonality in the data. Mean oocyst concentrations on hay paddock forages were likely to increase with increased prevalence (Fig. 4). The regression line of oocyst concentration versus oocyst prevalence on hay paddock forages was significant and showed that concentration increased 0.19 oocysts/g of forage for each 10% increase in prevalence. The regression relationship between oocyst concentration and prevalence in hay paddock soils was nearly identical to the relationship observed in pasture paddock soils (Fig. 5), but not statistically significant.

Pasture versus hay

Greater oocyst prevalences and concentrations in pasture soils and on pasture forages than in hay soils and on hay forages were expected if cattle were excreting oocysts in their manure. Others have found that adult cattle excrete oocysts in moderated concentrations relative to concentrations excreted by infected calves (Sturdee *et al.*, 2003). Infected adult cattle rarely exhibit clinical symptoms of cryptosporidiosis. Cattle

were not tested for *Cryptosporidium* infection in this study. Cattle spent much more time throughout the year in the pasture paddocks than in the hay paddocks, thus excreting more manure in the pasture paddocks.

Relatively high oocyst prevalences and concentrations in the hay paddocks were surprising since cattle were rarely present there. Wild animals might have been one source of oocysts throughout the year. Effects of wild animals on the prevalence and concentrations of *Cryptosporidium* oocysts in the soils and on the forage are unknown. Meadow voles (*Microtus pennsylvanicus*), deer mice (*Peromyscus maniculatus*), raccoons (*Procyon lotor*), and white-tailed deer (*Odocoileus virginianus*) are common in the area and all are known sources of *Cryptosporidium* spp. oocysts (Feng *et al.*, 2007). Wild animals had unrestricted access to all paddocks and could have easily transported oocysts between paddocks. Sturdee *et al.* (2003) suggested that wild animals chronically infected with *Cryptosporidium* could serve as a perpetual source of infection for domestic livestock.

Regardless of effects of wild animals, higher oocyst prevalences and concentrations in pasture indicate that cattle were a source of oocysts. However, oocyst concentrations in cattle feces in this study were not assessed. Other studies have found that cattle feces are a source of oocysts (Scott *et al.*, 1994; Atwill *et al.*, 1999, 2006). The cycle of oocyst ingestion, infection, and oocyst excretion is exacerbated in pasture by higher prevalences and concentrations. Further, as the chance of ingestion (prevalence on forage) increases, numbers of oocysts ingested increases because of higher oocyst concentrations. The expected number of oocysts ingested per day can be estimated by the equation

$$E(O_i) = P \times O_c \times F, \quad (1)$$

where $E(O_i)$ = expected number of oocysts ingested per day, P = proportion of positive samples (prevalence), O_c = mean number of oocysts/g of forage in samples that were positive for oocysts, and F = grams of forage ingested per day. If a 300 kg steer is considered to eat 3% of its weight in forage dry matter (9000 g)/day and the forage is 15% dry matter, then F equals 60,000 g/day. Substitution of the annual mean oocyst prevalences and concentrations (pasture: 0.52 and 1.44; hay: 0.41 and 1.00, respectively) into Equation 1 gives an expected mean daily oocyst ingestion per steer of 44,928 in the pasture paddock and 24,600 in the hay paddock.

Expected mean daily ingestion of oocysts can be estimated for prevalences using the linear trends shown in Figure 4. Substitution of prevalences from 0% to 70% for P and the associated oocyst concentration for O_c in Equation 1 gives the $E(O_i)$ as shown in Figure 6. Higher concentrations of oocysts on pasture paddock forage versus hay paddock forage causes the differences in expected mean daily ingestion between pasture and hay to increase with increasing prevalence. This exercise is only illustrative of potential differences between the two paddock managements. A great deal of uncertainty is associated with the relationship. First, the scatter around the lines in Figure 4, especially in the pasture paddocks, lends some uncertainty in the input values even though the slopes of both lines were significant. Additionally, Equation 1 is dependent on amount of forage ingested, which is determined by several factors, including animal size, forage dry matter content, and nutritional value of the forage. It is also impor-

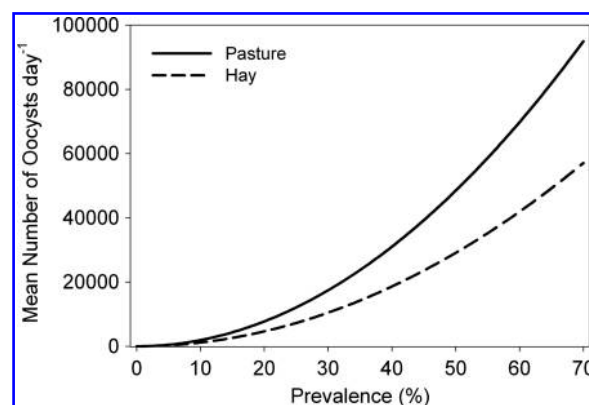


FIG. 6. Expected mean daily number of oocysts ingested with forage versus prevalence of oocysts on forage in pasture and hay paddocks. Calculations based on a 300 kg steer ingesting 60,000 g of forage per day.

tant to consider infectivity of oocysts. Oocysts are expected to lose infectivity over time, but such factors as temperature, desiccation, and ultraviolet radiation have all been shown to affect longevity of oocyst infectivity (Robertson *et al.* 1992; Olson *et al.*, 1999; Brookes *et al.*, 2004; Li *et al.*, 2005). Walker *et al.* (1998) suggested that vegetated soil protects oocysts from effects of temperature extremes and desiccation. Nevertheless, the data show that oocyst ingestion will be greater in pasture paddocks than in hay paddocks since prevalences and concentrations on the pasture forage is greater.

Water quality management challenges

Cryptosporidium is ubiquitous in surface water (Rose, 1997) and has been found in high concentrations in an Appalachian karst spring draining grazing lands (Boyer and Kuczynska, 2003). Samples drawn from a well in a shallow karst aquifer on this research site were found to contain *Cryptosporidium* oocysts (unpublished data). Protection from waterborne disease is the ultimate goal of landscape and water quality management for control of opportunities for *Cryptosporidium* oocysts to enter drinking water supplies.

Prevention of infection of livestock might be an effective means to prevent oocysts from reaching water supplies. Wild animals appear to be a source of environmental loading of oocysts on pasture. Sturdee *et al.* (2003) recommended that maintenance of good animal health might reduce severity of infection of grazing livestock and subsequent environmental loading of oocysts. Once oocysts are in the environment, management decisions and practices for reduction of opportunities for oocysts to reach water supplies are needed.

Reduction of surface runoff and exclusion of grazing animals from runoff source areas will reduce opportunities for oocysts to be transported to surface streams. Vegetative filter strips have often been found effective in reducing runoff and pathogen transport (Trask *et al.*, 2004). Deep infiltration of water might be counterproductive because oocysts have been found to survive for long times in temperate climate agricultural soils (Jenkins *et al.*, 2002). Oocysts have been found to leach through macroporous soils (Darnault *et al.*, 2004; Harter *et al.*, 2008; Boyer *et al.*, 2009) and remain infective for ex-

tended time (Boyer *et al.*, 2009). Management practices that leave vegetation short enough to allow radiant energy to reach the soil surface might produce enough surface heating and allow enough penetration of ultraviolet radiation to inactivate oocysts. Trevisan *et al.* (2002) suggested that canopy management of hay meadows might be an effective means for controlling fecal bacteria. Assuming that most oocysts on forage are located on lower levels of the forage, livestock would ingest fewer oocysts from short forage if they are moved to other pasture with taller forage. Short rotations might be one way to reduce the number of oocysts ingested, thereby reducing infection and subsequent environmental contamination.

On the basis of the high prevalences and concentrations of oocysts in the infrequently, lightly grazed hay paddocks, it appears that wild animals are an important source of oocysts in grazing systems. However, there is the possibility that oocysts are just persisting and reinfected cattle are maintaining the supply of oocysts. Control of wild animals, especially small animals, would be difficult if not nearly impossible. Multidisciplinary studies that include wildlife experts are needed to devise management strategies that lessen the role of wild animals in the dissemination and perpetuation of existence of oocysts in grazing systems. Runoff from grazed paddocks might have transported some oocysts onto hay paddocks, but previous infiltration studies (unpublished data) showed that infiltration rates on the site were high and runoff was not expected to be a problem. The farm-to-fork concept for protecting food safety targets all aspects of food production with control measures starting at the farm and ultimately ending at the consumer's fork or drinking cup (Callaway and Oliver, 2009). Identification of the causes of foodborne and waterborne contamination is needed to effectively manage associated risks (Jacob and Powell, 2009). Control of *Cryptosporidium* oocysts entering the environment at the farm level provides opportunities for reducing treatment costs and reducing public safety risks associated with food and water supplies.

Conclusions

Cryptosporidium oocysts were ubiquitous in surface soils and on forage in grazed paddocks as well as, infrequently, lightly grazed paddocks used for hay production. Prevalences and concentrations of oocysts were generally greater in the soils and on the forage of the pasture paddocks than the hay paddocks. Lack of seasonality in the prevalences of *Cryptosporidium* oocysts in soil suggests that oocysts are persistent. Persistence of *Cryptosporidium* oocysts in the hay paddock soils suggested that wildlife might be an important source and transport vector of *Cryptosporidium* oocysts in the area studied. High prevalence and concentration of *Cryptosporidium* oocysts on forage might be a source of infection to grazing livestock ingesting contaminated forage. Canopy management, short-cycle rotational grazing, and control of wildlife are potential strategies for reduction of *Cryptosporidium* oocysts in pasture and lessening the risk of contamination of water supplies, but further studies are needed before recommendations can be made. This study was limited to one specific geographical location with a single geology. Further studies across other locations and geologies are needed before broad recommendations can be made for pasture manage-

ment systems that reduce opportunities for *Cryptosporidium* oocysts to contaminate water supplies.

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Disclosure Statement

No competing financial interests exist.

References

- Atwill ER, Johnson EM, and Pereira MDGC. Association of herd composition, stocking rate, and duration of calving season with fecal shedding of *Cryptosporidium parvum* oocysts in beef herds. *J Am Vet Med Assoc* 1999;215:1833–1838.
- Atwill ER, Pereira MDGC, Alonso LH, Elmi C, Epperson WB, Smith R, Riggs W, Carpenter LV, Dargatz DA, and Hoar B. Environmental load of *Cryptosporidium parvum* oocysts from cattle manure in feedlots from the central and western United States. *J Environ Qual* 2006;35:200–206.
- Barwick RS, Mohammed HO, White ME, and Bryant RB. Prevalence of *Giardia* spp. and *Cryptosporidium* spp. on dairy farms in southeastern New York state. *Prev Vet Med* 2003a; 59:1–11.
- Barwick RS, Mohammed HO, White ME, and Bryant RB. Factors associated with the likelihood of *Giardia* spp. and *Cryptosporidium* spp. in soil from dairy farms. *J Dairy Sci* 2003b;86:784–791.
- Bodley-Tickell AT, Kitchen SE, and Sturdee AP. Occurrence of *Cryptosporidium* in agricultural surface waters during an annual farming cycle in lowland UK. *Water Res* 2002;36:1880–1886.
- Boyer DG and Belesky DP. Interactions of slope and canopy of herbage of three herbage species on transport of faecal indicator bacteria by rain splash. *Grass Forage Sci* 2009;64:432–442.
- Boyer DG and Kuczynska E. Storm and seasonal distributions of fecal coliforms and *Cryptosporidium* in a spring. *J Am Water Resour Assoc* 2003;39:1449–1556.
- Boyer DG, Kuczynska E, and Fayer R. Transport, fate, and infectivity of *Cryptosporidium parvum* oocysts released from manure and leached through macroporous soil. *Environ Geol* 2009;58:1011–1019.
- Brookes JD, Antenucci J, Hipsey M, Burch MD, Ashbolt NJ, and Ferguson C. Fate and transport of pathogens in lakes and reservoirs. *Environ Int* 2004;30:741–759.
- Callaway TR and Oliver SP. On-farm strategies to reduce foodborne pathogen contamination. *Foodborne Pathog Dis* 2009;6:753.
- [CDC] Centers for Disease Control and Prevention. Assessing the Public Health Threat Associated with Waterborne *Cryptosporidiosis*: Report of a Workshop. *Morb Mortal Wkly Rep* 1995;44:1–19. Available at www.cdc.gov/mmwr/PDF/rr/rr4406.pdf, accessed October 22, 2009. (Online.)

- Darnault CJG, Steenhuis TS, Garnier P, Kim Y-J, Jenkins MB, Ghiorse WC, Baveye PC, and Parlange J-Y. Preferential flow and transport of *Cryptosporidium parvum* oocysts through the vadose zone: experiments and modeling. *Vadose Zone J* 2004;3:262–270.
- Davies CM, Ferguson CM, Kaucner C, Krogh M, Altavilla N, Deere DA, and Ashbolt NJ. Dispersion and transport of *Cryptosporidium* oocysts from fecal pats under simulated rainfall events. *Appl Environ Microbiol* 2004;70:1151–1159.
- DuPont HL, Chappell CL, Sterling CR, Okhuysen PC, Rose JB, and Jakubowski W. The infectivity of *Cryptosporidium parvum* in healthy volunteers. *N Engl J Med* 1995;332:855–859.
- Fayer R, Trout JM, Graczyk TK, and Lewis EJ. Prevalence of *Cryptosporidium*, *Giardia* and *Eimeria* infections in post-weaned and adult cattle on three Maryland farms. *Vet Parasitol* 2000;93:103–112.
- Feng Y, Alderisio KA, Yang W, Blancero LA, Kuhne WG, Nardeski CA, Reid M, and Xiao L. *Cryptosporidium* genotypes in wildlife from a New York watershed. *Appl Environ Microbiol* 2007;73:6475–6483.
- Hamilton PD, Gale P, and Pollard SJT. A commentary on recent water safety initiatives in the context of water utility risk management. *Environ Int* 2006;32:958–966.
- Harter T, Atwill ER, Hou L, Karle BM, and Tate KW. Developing risk models of *Cryptosporidium* transport in soils from vegetated, tilted soilbox experiments. *J Environ Qual* 2008;37:245–258.
- Hubbard RK, Newton GL, and Hill GM. Water quality and the grazing animal. *J Anim Sci* 2004;82(supp E):255–263.
- Jacob CJ and Powell DA. Where does foodborne illness happen—in the home, at foodservice, or elsewhere—and does it matter? *Foodborne Pathog Dis* 2009;6:1121–1123.
- Jenkins MB, Bowman DD, Fogarty EA, and Ghiorse WC. *Cryptosporidium parvum* oocyst inactivation in three soil types at various temperatures and water potentials. *Soil Biol Biochem* 2002;34:1101–1109.
- Karanis P, Kourenti C, and Smith H. Waterborne transmission of protozoan parasites: a worldwide review of outbreaks and lessons learnt. *J Water Health* 2007;5:1–38.
- Kemp JS, Wright SE, and Bukhari Z. On farm detection of *Cryptosporidium parvum* in cattle, calves and environmental samples. In: *Protozoan Parasites and Water*, Special Publication, Volume 168. Betts WB, Casemore D, Fricker C, Smith H, and Watkins J (eds.). Cambridge: Royal Society of Chemistry, 1995, pp. 154–157.
- King J and Monis PT. Critical processes affecting *Cryptosporidium* oocyst survival in the environment. *Parasitology* 2006;134:309–323.
- Kuczynska E and Shelton DR. Method for detection and enumeration of *Cryptosporidium parvum* oocysts in feces, manures, and soils. *Appl Environ Microbiol* 1999;65:2820–2826.
- Kuczynska E, Shelton DR, and Pachepsky Y. Effect of bovine manure on *Cryptosporidium parvum* oocysts attachment to soil. *Appl Environ Microbiol* 2005;71:6394–6397.
- LeChevallier MW, Norton WD, and Lee RG. Occurrence of *Giardia* and *Cryptosporidium* spp. in surface water supplies. *Appl Environ Microbiol* 1991;57:2610–2616.
- Li X, Atwill ER, Dunbar LA, Jones T, Hook J, and Tate KW. Seasonal temperature fluctuations induces rapid inactivation of *Cryptosporidium parvum*. *Environ Sci Technol* 2005;39:4484–4489.
- Mawdsley JL, Bardgett RD, Merry RJ, Pain BF, and Theodorou MK. Pathogens in livestock waste, their potential for movement through soil and environmental pollution. *Appl Soil Ecol* 1995;2:1–15.
- Olson ME, Goh J, Phillips M, Guselle N, and McAllister TA. *Giardia* cyst and *Cryptosporidium* oocyst survival in water, soil, and cattle feces. *J Environ Qual* 1999;28:1991–1996.
- Ong C, Moorehead W, Ross A, and Isaac-Renton J. Studies of *Giardia* spp. and *Cryptosporidium* spp. in two adjacent watersheds. *Appl Environ Microbiol* 1996;62:2798–2805.
- Paul PA, El-Allaf SM, Lipps PE, and Madden LV. Rain splash dispersal of *Gibberella zeae* within wheat canopies in Ohio. *Phytopathology* 2002;94:1342–1349.
- Robertson LJ, Campbell AT, and Smith HV. Survival of *Cryptosporidium parvum* oocysts under various environmental pressures. *Appl Environ Microbiol* 1992;58:3494–3500.
- Rose JB. Environmental ecology of *Cryptosporidium* and public health implications. *Annu Rev Public Health* 1997;18:135–161.
- Schijven JF, Bradford SA, and Yang S. Release of *Cryptosporidium* and *Giardia* from dairy cattle manure: physical factors. *J Environ Qual* 2004;33:1499–1508.
- Scott CA, Smith HV, and Gibbs HA. Excretion of *Cryptosporidium parvum* oocysts by a herd of beef suckler cows. *Vet Rec* 1994;134:172.
- States S, Stadterman K, Ammon L, Vogel P, Baldizar J, Wright D, Conley L, and Sykora J. Protozoa in river water: sources, occurrence, and treatment. *J Am Water Works Assoc* 1997;89:74–83.
- Sturdee AP, Bodley-Tickell AT, Archer A, and Chalmers RM. Long-term study of *Cryptosporidium* prevalence on a lowland farm in the United Kingdom. *Vet Parasitol* 2003;116:97–103.
- Sturdee A, Foster I, Bodley-Tickell AT, and Archer A. Water quality and *Cryptosporidium* distribution in an upland water supply catchment, Cumbria, UK. *Hydrol Process* 2007;21:873–875.
- Trask JR, Kalita PK, Kuhlenschmidt MS, Smith RD, and Funk TL. Overland and near-surface transport of *Cryptosporidium parvum* from vegetated and nonvegetated surfaces. *J Environ Qual* 2004;33:984–993.
- Trevisan D, Vansteelant JY, and Dorioz JM. Survival and leaching of fecal bacteria after slurry spreading on mountain hay meadows: consequences for the management of water contamination risk. *Water Res* 2002;36:275–283.
- Walker MJ, Montemagno CD, and Jenkins MB. Source water assessment and nonpoint sources of acutely toxic contaminants: a review of research related to survival and transport of *Cryptosporidium parvum*. *Water Resour Res* 1998;34:3383–3392.

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